

THROMBOXAN A₂ RECEPTOR (TPR α): A POTENTIAL HUMAN DRUG TARGET

Biochemistry
Judy Conmey
Senior Sophister

The human thromboxane A₂ receptor (TPR α), a member of the superfamily of G protein-coupled receptors, is involved in platelet activation and vasoconstriction in response to binding of thromboxane A₂ (TXA₂). However, over-activation of TPR α can lead to the development of thrombotic cardiovascular events such as heart attacks and strokes. Currently the prevention of secondary cardiovascular events is carried out by the administration of aspirin. The TPR α is an important target for drug development due to its role in platelet aggregation and vasoconstriction. This review will focus the potential of TPR α as a human drug target.

Introduction

Thromboxane A₂ is a prostanoid that activates vasoconstriction and platelet aggregation through interaction with the thromboxane A₂ receptor (TPR α). The official name of this receptor is the thromboxane A₂ receptor (Magrane M. and the UniProt consortium 2011), but it is also known as the thromboxane/prostaglandin H₂ (PGH₂) receptor or the prostanoid TP receptor (Coleman *et al.* 1994, Ting *et al.* 2012). For the purpose of this review, thromboxane A₂ and the thromboxane A₂ receptor will be referred to as TXA₂ and TPR α , respectively.

TPR α is a G-protein coupled receptor (GPCR) expressed in human blood platelets and plays a vital role in platelet activation through coupling to the G-protein G_q (among others) (Ting *et al.* 2012). Although it binds both TXA₂ and PGH₂, the ligand with greater affinity for this receptor is TXA₂ (Devillier & Bessard 1997).

Activation of TPR α by TXA₂ triggers cellular signalling responses which result in platelet adhesion, vesicle trafficking, and changes in the cytoskeleton (Ting *et al.* 2012), all essential for haemostasis. However, if TPR α is over-activated serious thrombotic cardiovascular diseases may develop such as acute myocardial infarction, hypertension and unstable angina (Halushka & Halushka 2002).

Aspirin is currently used to inhibit platelet TXA₂ synthesis as a means of treating a variety of thrombotic cardiovascular diseases, such as acute myocardial infarction, unstable angina and secondary prevention of myocardial infarction (Morinelli & Halushka 1991, Mousa 1999). Aspirin inhibits cyclooxygenases (COXs), key enzymes involved in the biosynthesis of TXA₂ (Fig. 1) (Mousa 1999).

However, as mentioned, TXA₂ is not the only endogenous ligand for TPR α . Therefore aspirin inhibits just one of the many pathways that can result in platelet aggregation (Mousa 1999). Aspirin, despite its success in the treatment of secondary acute myocardial infarction, has efficacy issues. There is a proportion of the population who may experience “aspirin resistance”, with explanations for this resistance still inconclusive (Halushka & Halushka 2002, Gasparyan *et al.* 2008). Furthermore, low doses of aspirin carry a risk of increased upper gastrointestinal side effects, such as bleeding and ulcers, which could become life-threatening (Cryer & Feldman 1999). Despite Bousser *et al.* (2011, p. 2013) concluding that aspirin is still “the gold standard anti-platelet drug for secondary stroke prevention in view of its efficacy, tolerance, and cost”, it is responsible for a high rate of people being hospitalised due to a bad reaction to the drug (Pirmohamed *et al.* 2004).

Thus, the development of a new anti-platelet drug is necessary for the treatment of numerous diseases where platelet activity is promoted. The three-dimensional structure of TPR α at atomic resolution would aid the design of effective anti-platelet agents by providing a molecular insight into how the receptor functions.

This review aims analyse what is known about TPR α in current literature and highlight some considerations when targeting TPR α .

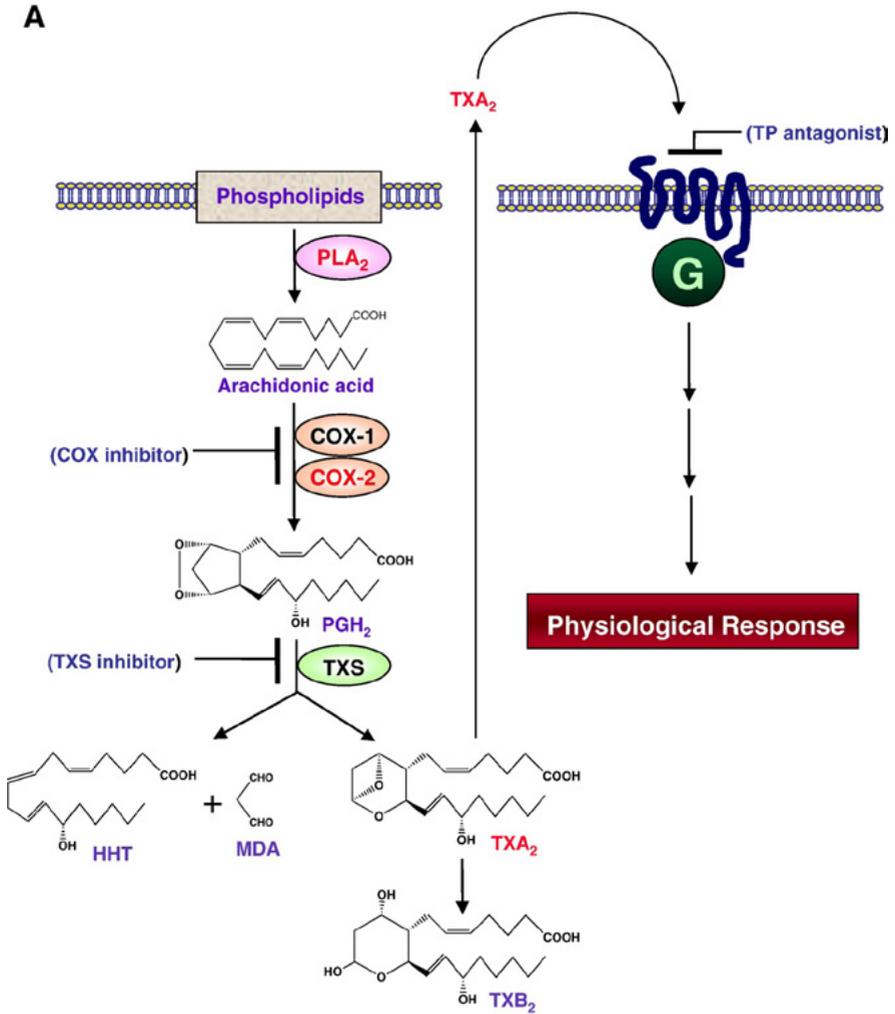


Figure 1: The human TXA₂ biosynthetic pathway. The chronological steps of thromboxane A₂ (TXA₂) synthesis and the stages where drugs act, affecting TXA₂ function, are shown. TXA₂ is synthesised from arachidonic acid (AA) which is released from the membrane by phospholipase A₂ (PLA₂). Cyclooxygenases (COX-1 and COX-2) convert AA into important biological mediators known as prostanoids, the first being prostaglandin H₂ (PGH₂), a precursor of thromboxane A₂ (TXA₂). PGH₂ is converted to TXA₂, 12-L-hydroxy-5,8,10-heptadecatrienoic acid (HHT) and malondialdehyde (MDA) by thromboxane synthase (TXS). Thromboxane B₂ (TXB₂) is a product of TXA₂ hydrolysis that is biologically inactive and removed from the body. (Nakahata 2008)

1. Functions of Thromboxane A₂ and Thromboxane A₂ Receptor

1.1. Biosynthesis of TXA₂

TXA₂ is an unstable product of the cyclooxygenase (COX) mediated metabolism of arachidonic acid (AA), catalysed by phospholipase A₂ (PLA₂). COXs synthesise the endoperoxide prostaglandin, prostaglandin H₂ (PGH₂), from AA which is in turn converted to TXA₂ by thromboxane synthase (TXS). TXA₂ has a short half-life of about 30 seconds (Fig. 1) before being converted into the inactive TXB₂ (Hamberg *et al.* 1975). Thus, TXA₂ acts on neighbouring cells in close proximity to the cell where it is produced in an autocrine or paracrine manner (Nakahata 2008).

1.2. TXA₂ and the TPR α Signalling Pathway

Binding of a TXA₂ to TPR α elicits several molecular responses which, overall, result in the elevation of intracellular Ca²⁺ levels which is the key to platelet activation (Fig. 2) (Brass *et al.* 1997).

Several G-protein isoforms are stimulated by TPR α upon ligand binding. In 2000, Perry V. Halushka outlined that there are at least nine G proteins which couple to TPR α ; G α_q , G α_{12} , G α_s , G α_{11} , G α_{12} , G α_{13} , G α_{15} , G α_{16} and G α_h . The G α_q isoform is of interest as TPR α -mediated platelet aggregation primarily associated with G α_q (Offermanns *et al.* 1994). The G α_q family activates Phospholipase C beta (PLC- β) which cleaves the phospholipid phosphatidylinositol 4,5-bisphosphate (PIP₂) to form second messengers inositol 1,4,5-triphosphate (IP₃) and diacylglycerol (DAG) (Ting *et al.* 2012). IP₃ releases intracellular calcium stores and DAG activates protein kinase C (Fig. 3) (Berridge & Irvine 1984, Kikkawa *et al.* 1986). Overall, these responses lead to platelet activation (Huang *et al.* 2004).

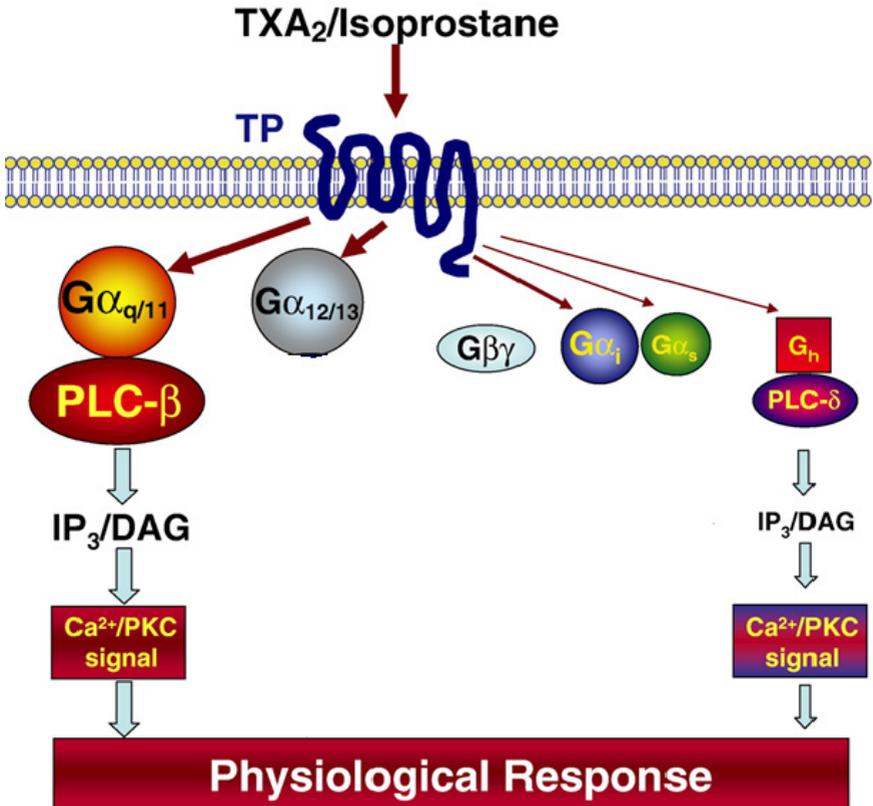


Figure 2: G protein coupling of thromboxane A₂ receptor and signal transduction
 When ligand bound, TPR_α can couple to several different G proteins. When TPR_α is bound to G_{α11/13}, phospholipase C-β (PLC-β) is activated, which converts phosphatidylinositol 4,5-bisphosphate (PIP₂) into 2nd messengers inositol 1,4,5-trisphosphate (IP₃) and diacylglycerol (DAG). IP₃ releases intracellular calcium stores and DAG activates protein kinase C (Berridge & Irvine 1984, Kikkawa et al. 1986). (adapted from Nakahata 2008).

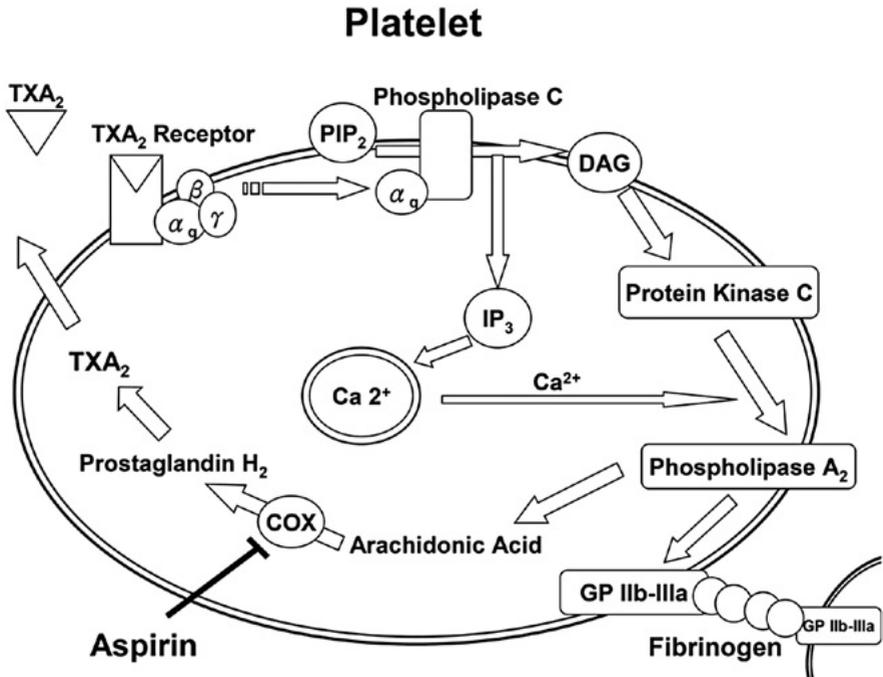


Figure 3: TXA₂ and TPRα intracellular signalling. Gq protein activates phospholipase C, which hydrolyses phosphatidylinositol 4,5-bisphosphate (PIP₂) to produce inositol 1,4,5-triphosphate (IP₃) and diacylglycerol (DAG). IP₃ elevates the cytosolic free Ca²⁺ concentration by inducing vesicular release of Ca²⁺ into cytosol. DAG activates protein kinase C and activates phospholipase A₂ stimulation of phospholipase A₂ is associated with the activation of glycoprotein IIb/IIIa complex, leading fibrinogen cross-linkage by binding to glycoprotein IIb/IIIa receptors present on each platelet (Jeng et al. 1998). Activation of PLA₂ also amplifies the signal transduction message by the generation of more TXA₂ from arachidonic acid by cyclooxygenase (COX). Aspirin inhibits COXs, down-regulating TXA₂ and thus platelet aggregations and clot formation. (Shiau et al. 2012)

1.3. Physiological/Pathophysiological role of TXA₂ and TXA₂ Receptors in cardiovascular diseases

Platelet activation is a significant physiological response when TXA₂ binds TPRα. Platelet activation leads to alterations in platelet shape, aggregation and secretion. This in turn stimulates thrombus formation (Ally & Horrobin 1980, Dorn & DeJesus 1991, Spurney et al. 1991, Katugampola & Davenport 2001). TXA₂-induced platelet aggregation and vasoconstriction can lead to acute myocardial

infarction (Martin 1984, Hiraku *et al.* 1986, Dorn *et al.* 1990, Katugampola & Davenport 2001). These disease states are currently treated by TXA₂ synthesis inhibitors but not TPR α antagonists. TPR α antagonists could in theory be an effective means of treatment, but to date development of this class of drugs has been unsuccessful. TXA₂ is a ligand well-known to exert its actions through TPR α , but it is not the only endogenous ligand to do so. There is current evidence that suggests isoprostane 8-iso-prostaglandin F₂ α (8-iso PGF₂ α) is another TPR α stimulant (Montuschi *et al.* 2004). Isoprostanes are peroxidative products of AAs, and are more stable when compared to TXA₂ (Montuschi *et al.* 2004). Therefore, TXA₂ may not be the primary ligand responsible for the cardiovascular diseases associated with TPR α -mediated signalling.

2. Physiological/Pathophysiological Effects of Targeting TPR α -Mediated Signalling

2.1. Marketed Drugs which Target TPR α -mediated Signalling

An effective antagonist targeting TPR α -mediated signalling has not been synthesised to date due, in part, to the fact not enough is known about what happens structurally when TPR α becomes activated. If the crystal structure was determined then it is possible that an effective drug could be synthesised based on the structure of the receptor. As the antagonist would be specific for the TPR α , it should not, in theory, have any serious side effects. To further qualify it as effective and successful in clinical trials, the proposed TPR α drug should be suitable for oral or intravenous administration, like that of aspirin. Also, the length of time of which the drug acts in the body should be adequate to therapeutically treat the disease state (Mousa 1999) i.e. it must be stable and not breakdown into a potentially dangerous by-product *in-vivo*.

The class of drugs which target TXA₂ production itself have been very successful to date (aspirin and dazoxiban). Aspirin and non-steroidal anti-inflammatory drugs (NSAIDS) are cyclooxygenase inhibitors which inhibit TXA₂ at the early stages of biosynthesis.

These inhibitors stop TXA_2 production by blocking arachidonic acid conversion to PGH_2 (Fig. 1). There are three cyclooxygenase (COX) isoenzymes identified; COX-1, COX-2 and COX-3. Platelets express COX-1 which can be inhibited non-selectively (i.e. by COX inhibitors which target both COX-1 and COX-2). Aspirin inhibits COX-1 by irreversibly acetylating a serine residue in the catalytic channel (Roth *et al.* 1977). At higher concentrations, COX-2 may also be inhibited by aspirin (Cipollone *et al.* 1997). This action leads to long-term down-regulation of TXA_2 synthesis in platelets as they don't possess a nucleus and cannot make their own COX-1.

Low dose aspirin remains the best standard of secondary prevention after myocardial infarction or stroke and more than 80% of such patients take aspirin regularly (Campbell *et al.* 2007). Despite the success of this class of drugs, none of the current anti-platelet agents available to the public meet all of the requirements outlined, as their lack of selectivity result in undesirable side-effects (Mousa 1999).

A negative side-effect of the non-selectivity of aspirin is that it may also inhibit PGI_2 synthesis, along with several other prostaglandins (Fig. 4) (Coccheri 2010). PGI_2 is a lipid essential for the maintenance of the endothelium and its inhibition can lead to deleterious side-effects such as disruption of the gastrointestinal lining. The ability of aspirin to inhibit PGI_2 production is believed to be relevant to explaining why some people are sensitive to aspirin while others are resistant (Pusch *et al.* 2008, Coccheri 2010). It is for these reasons that research efforts have shifted to focus on other targets in the signalling pathway.

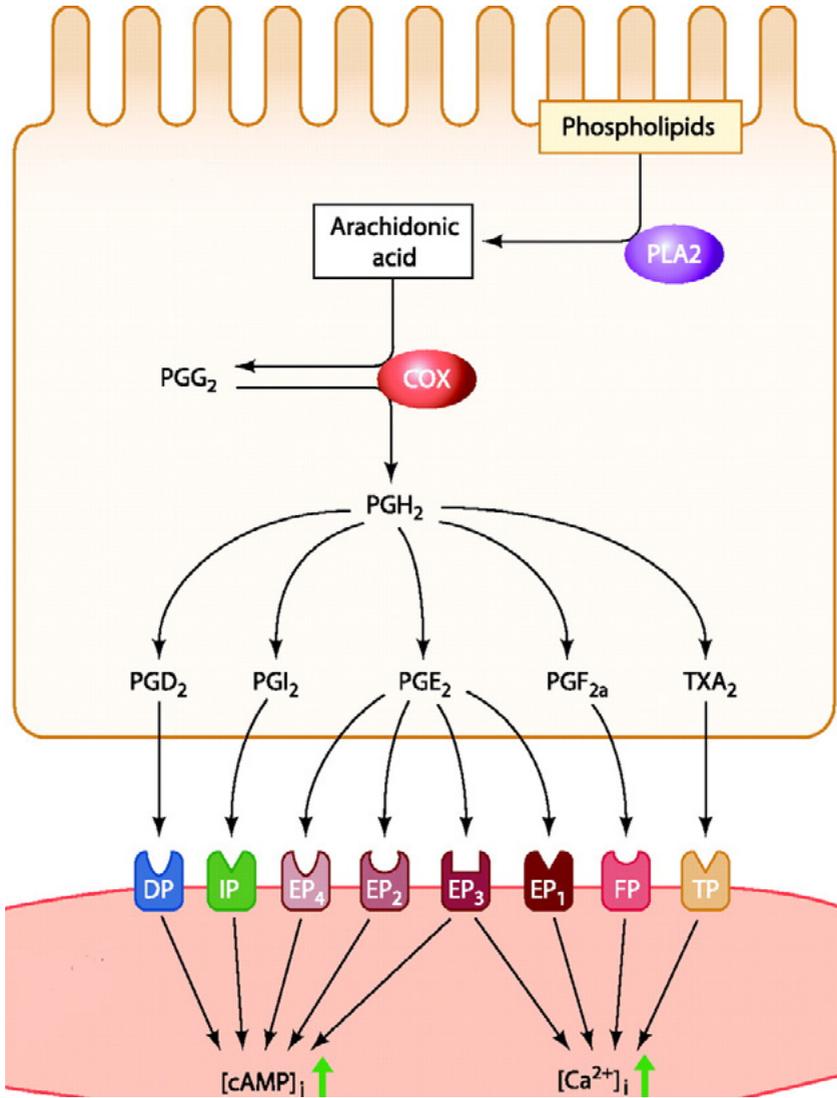


Figure 4: The cyclooxygenase (COX) and prostanoid biosynthetic and signalling pathways. Arachidonic acid (AA) is released by phospholipase A₂ (PLA₂) and is acted on by COX enzymes and synthase enzymes (prostaglandin synthases and thromboxane synthase). These enzymes synthesise their respective prostaglandins and they move out of the cell through prostaglandin transporter (PGT). When aspirin blocks COX-1, it invariably inhibits the all of these signalling cascades. (adapted from Ruan et al. 2011)

2.2. Thromboxane Synthase (TXS) Inhibitors

Thromboxane synthase (TXS) inhibitors were developed to block conversion of PGH_2 to TXA_2 . These drugs proved successful as they reduced TXA_2 synthesis in platelets and improved TXA_2 -related pathophysiological conditions (Dogne *et al.* 2005). Many drugs of this class were designed but unfortunately failed at clinical trials due to the accumulation of PGH_2 , which increases platelet activity by acting as an agonist for $\text{TPR}\alpha$ (Ting *et al.* 2012). If a drug was produced which targeted the downstream receptor as opposed to the TXA_2 synthesis itself, it would reduce the effects of TXA_2 but PGI_2 production would be unaffected.

The added advantage of direct antagonism of $\text{TPR}\alpha$ over TXA_2 synthesis inhibition is that such drugs would block other ligands which have agonistic effects on the receptor, such as endoperoxides and isoprostanes (Gresele *et al.* 1987, Fiddler & Lumley 1990). There is evidence to show that $\text{TPR}\alpha$ mediates signalling through multiple effectors, including phospholipase C (PLC), some small guanosine triphosphate hydrolases (GTPases) and adenylyl cyclase (AC) (Ting *et al.* 2012). Furthermore, $\text{TPR}\alpha$ can recognise endogenous ligands other than eicosanoid thromboxane A_2 (TXA_2), such as the endoperoxide PGG_2 isoprostanes (Khasawneh *et al.* 2008). It will be important to considering the implications of the downstream pathways of these ligands when antagonising/blocking receptor.

Unfortunately, little progress has been made with successfully targeting these receptors through development of drugs due to problems with efficacy and toxicity. Most antagonists synthesised have not passed phase I/II clinical trials (Ting *et al.* 2012). In 2011, Bousser *et al.* studied terutroban. They interpreted no safety or efficacy advantages with terutroban versus aspirin. However, this study is inconclusive as it was prematurely stopped by the Data Monitoring Committee as it didn't meet the predefined criteria for non-inferiority (Bousser *et al.* 2011). Non-inferiority trials are intended to show that the effect of a new treatment is not worse than that of an active control, aspirin in this case, by more than a specified margin (Snapinn 2000). Therefore, research in developing drugs to target $\text{TPR}\alpha$ would benefit from the high resolution structure

determination.

3. Structural Features of TPR α

3.1. G-Protein Coupled Receptors

TPR α is one of five prostanoid receptor classifications recognised (Coleman *et al.* 1994). TPR α is a member of a large superfamily of transmembrane proteins, GPCRs, that are implicit in various cellular signals (Moreira 2014).

The GPCR superfamily is comprised of over 800 members, making it one of the largest protein families in the human genome. Not surprisingly it is associated with numerous human diseases and 30% of marketed drugs target GPCRs (Fredriksson *et al.* 2003, Hopkins & Groom 2002). Phylogenetic analysis has identified five main families that classify human GPCRs; rhodopsin-like (672 members), secretin (15 members), glutamate (22 members), adhesion (33 members) and frizzled-taste 2 (36 members) (Moreira 2014). Of these, TPR α belongs to the rhodopsin-like, also known as sub-family A.

The topology of sub-family A GPCRs comprises seven transmembrane α helices (TM1-7) in addition to an extracellular N-terminus and an intracellular C-terminus (Moreira 2014). These helices are connected by three extracellular (ECL) and three intracellular (ICL) loops.

3.2. Coupling of TPR α to G proteins

GPCR signalling involves the coupling of the activated receptor to a heterotrimeric GTP-binding protein (G-protein) (Jastrzebska *et al.* 2010). In the case of TPR α , binding occurs to the G-protein G_q among others. TPR α can only be associated with one G-protein at a time and receptor recycling regulates the specificity of the G-protein (Ting *et al.* 2012). The G_q protein is divided into two functional units, the guanine nucleotide-binding G α subunit and the G $\beta\gamma$ dimer (Fig. 5). When activated, the functional units dissociate and these in turn regulate a large number of downstream effects through association with enzymes and effector proteins (Hamm 1998). As

these receptors are so influential with regards to signal transduction in the cell in which they are expressed, it is important to structurally characterise them. However, only recently was the first GPCR/G-protein complex structure characterised (Rasmussen *et al.* 2011). This was a significant breakthrough as this was the first high-resolution view of transmembrane signalling by a GPCR in an activated state (R^*) and in complex with its endogenous G-protein. The active state of a GPCR can be defined as that conformation that couples to and stabilises a nucleotide-free G protein (Rasmussen *et al.* 2011). Having a picture of the transmembrane protein in an active conformation provides crucial information of how ligand binding affects structure and thus enables design of better agonists/antagonists of the target protein.

Therefore, it would be beneficial to solve the structure not only of $TPR\alpha$, but also that of $TPR\alpha$ - G_q complex. If the $TPR\alpha$ - G_q complex structure was determined then one would have a picture of the $TPR\alpha$ in R^* state and it might be possible to study how $TPR\alpha$ responds to ligand binding and how it this changes the conformation of the receptor by comparison of the co-complex structure to that of the receptor on its own.

For Educational Use Only

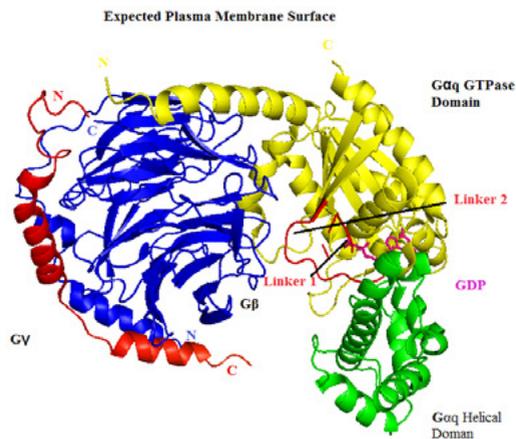


Figure 5: Structural representation of Heterotrimeric G_q -protein (PDB ID code 3AH8). Generated using Pymol. $G\alpha_{i/q}\beta\gamma$ heterotrimer orientated with respect to the plasma membrane. $G\alpha_{i/q}$ consists of the GTPase and the helical domains connected by two linker

regions, *Linker 1 and Switch I (Linker 2)* (Nishimura *et al.* 2010).

Conclusions

TPR α is not a “potential” drug target but it is in fact already a current human drug target for treatment of multiple thrombotic disorders. There has been a great deal of interest in regulating this receptor’s activity as an alternative to circumvent the side-effects observed with aspirin (Halushka & Halushka 2002). Aspirin’s side-effects are observed due to its lack of selectivity as it antagonizes both COX-1 and -2. Aspirin inhibits PGI₂ production, which is essential for maintenance of the endothelium which explains why patients who regularly take aspirin develop disruption of the gastrointestinal lining e.g. stomach ulcers. The next target in the pathway is thromboxane synthase (TXS), but inhibitors of this enzyme have been unsuccessful in clinical trials due to accumulation of PGH₂ which increases platelet activity (Ting *et al.* 2012). In theory, direct inhibition of downstream receptor itself, TPR α , presents many advantages as a drug target over the blocking of its upstream regulators. The advantage of an antagonist specific for TPR α is that all ligands which up-regulate the receptor would be blocked, not just TXA₂. Furthermore, PGI₂ production would be unaffected, thus limiting the possibility of side-effects, and PGH₂ accumulation would not be an issue. Unfortunately, the vast majority of compounds targeting TPR α failed to pass clinical trials or toxicity screenings as mentioned previously (Ting *et al.* 2012). The determination of the high resolution structure of TPR α may be a deciding factor to its success as a drug target as it will aid in rational, structure based drug design. While none of the antagonists developed for targeting TPR α to date have made it into clinical use, it may still prove a lucrative investment both intellectually and financially.

Acknowledgements

This work was adapted from a mini-review “Thromboxane A2 Receptor (TPR α): A Potential Human Drug Target” for course BI3015: LITERATURE SKILLS.

References

- ALLY, A. I. & HORROBIN, D. F. 1980. Thromboxane A2 in blood vessel walls and its physiological significance: relevance to thrombosis and hypertension. *Prostaglandins Med*, 4, 431-438.
- BERRIDGE, M. J. & IRVINE, R. F. 1984. Inositol trisphosphate, a novel second messenger in cellular signal transduction. *Nature*, 312, 315-21.
- BOUSSER, M. G., AMARENCO, P., CHAMORRO, A., FISHER, M., FORD, I., FOX, K. M., HENNERICI, M. G., MATTEL, H. P., ROTHWELL, P. M., DE CORDOUE, A. & FRATACCI, M. D. 2011. Terutroban versus aspirin in patients with cerebral ischaemic events (PERFORM): a randomised, double-blind, parallel-group trial. *Lancet*, 377, 2013-22.
- BRASS, L. F., MANNING, D. R., CICHOWSKI, K. & ABRAMS, C. S. 1997. Signaling through G proteins in platelets: to the integrins and beyond. *J Thromb Haemost*, 78, 581-9.
- CAMPBELL, C. L., SMYTH, S., MONTALESCOT, G. & STEINHUBL, S. R. 2007. Aspirin dose for the prevention of cardiovascular disease: a systematic review. *Jama*, 297, 2018-24.
- CIPOLLONE, F., PATRIGNANI, P., GRECO, A., PANARA, M. R., PADOVANO, R., CUCCURULLO, F., PATRONO, C., REBUZZI, A. G., LIUZZO, G., QUARANTA, G. & MASERI, A. 1997. Differential suppression of thromboxane biosynthesis by indobufen and aspirin in patients with unstable angina. *Circulation*, 96, 1109-16.
- COCCHERI, S. 2010. Antiplatelet drugs--do we need new options? With a reappraisal of direct thromboxane inhibitors. *Drugs*, 70, 887-908.
- COLEMAN, R. A., SMITH, W. L. & NARUMIYA, S. 1994. International Union of Pharmacology classification of prostanoid receptors: properties, distribution, and structure of the receptors and their subtypes. *Pharmacol Rev*, 46, 205-29.
- CRYER, B. & FELDMAN, M. 1999. Effects of very low dose daily, long-term aspirin therapy on gastric, duodenal, and rectal prostaglandin levels and on mucosal injury in healthy humans. *Gastroenterology*, 117, 17-25.
- DEVILLIER, P. & BESSARD, G. 1997. Thromboxane A2 and related prostaglandins in airways. *Fundam Clin Pharmacol*, 11, 2-18.
- DOGNE, J. M., HANSON, J. & PRATICO, D. 2005. Thromboxane, prostacyclin and

- isoprostanes: therapeutic targets in atherogenesis. *Trends Pharmacol Sci*, 26, 639-44.
- DORN, G. W., 2ND & DEJESUS, A. 1991. Human platelet aggregation and shape change are coupled to separate thromboxane A₂-prostaglandin H₂ receptors. *Am J Physiol*, 260, H327-34.
- DORN, G. W., 2ND, LIEL, N., TRASK, J. L., MAIS, D. E., ASSEY, M. E. & HALUSHKA, P. V. 1990. Increased platelet thromboxane A₂/prostaglandin H₂ receptors in patients with acute myocardial infarction. *Circulation*, 81, 212-8.
- FIDDLER, G. I. & LUMLEY, P. 1990. Preliminary clinical studies with thromboxane synthase inhibitors and thromboxane receptor blockers. A review. *Circulation*, 81, 169-178.
- FREDRIKSSON, R., LAGERSTROM, M. C., LUNDIN, L. G. & SCHIOTH, H. B. 2003. The G-protein-coupled receptors in the human genome form five main families. Phylogenetic analysis, paralogon groups, and fingerprints. *Mol Pharmacol*, 63, 1256-1272.
- GASPARYAN, A. Y., WATSON, T. & LIP, G. Y. H. 2008. The Role of Aspirin in Cardiovascular Prevention: Implications of Aspirin Resistance. *Journal of the American College of Cardiology*, 51, 1829-1843.
- GRESELE, P., ARNOUT, J., DECKMYN, H., HUYBRECHTS, E., PIETERS, G. & VERMYLEN, J. 1987. Role of proaggregatory and antiaggregatory prostaglandins in hemostasis. Studies with combined thromboxane synthase inhibition and thromboxane receptor antagonism. *J Clin Invest*, 80, 1435-45.
- HALUSHKA, M. K. & HALUSHKA, P. V. 2002. Why are some individuals resistant to the cardioprotective effects of aspirin? Could it be thromboxane A₂? *Circulation*, 105, 1620-2.
- HAMBERG, M., SVENSSON, J. & SAMUELSSON, B. 1975. Thromboxanes: a new group of biologically active compounds derived from prostaglandin endoperoxides. *Proc Natl Acad Sci U S A*, 72, 2994-8.
- HAMM, H. E. 1998. The many faces of G protein signaling. *J Biol Chem*, 273, 669-72.
- HIRAKU, S., TANIGUCHI, K., WAKITANI, K., OMAWARI, N., KIRA, H., MIYAMOTO, T., OKEGAWA, T., KAWASAKI, A. & UJIIE, A. 1986. Pharmacological studies on the TXA₂ synthetase inhibitor (E)-3-[p-(1H-imidazol-1-ylmethyl)phenyl]-2-propenoic acid (OKY-046). *Jpn J Pharmacol*, 41, 393-401.
- HOPKINS, A. L. & GROOM, C. R. 2002. The druggable genome. *Nat Rev Drug Discov*, 1, 727-30.
- HUANG, J.-S., RAMAMURTHY, S. K., LIN, X. & LE BRETON, G. C. 2004. Cell signalling through thromboxane A₂ receptors. *Cellular Signalling*, 16, 521-533.

- JASTRZEBSKA, B., TSYBOVSKY, Y. & PALCZEWSKI, K. 2010. Complexes between photoactivated rhodopsin and transducin: progress and questions. *Biochem J* **428**, 1-10.
- JENG, J.-S., LEE, T.-K., CHANG, Y.-C., HUANG, Z.-S., NG, S.-K., CHEN, R.-C. & YIP, P.-K. 1998. Subtypes and case-fatality rates of stroke: A hospital-based stroke registry in Taiwan (SCAN-IV). *Journal of the Neurological Sciences* **156**, 220-226.
- KATUGAMPOLA, S. D. & DAVENPORT, A. P. 2001. Thromboxane receptor density is increased in human cardiovascular disease with evidence for inhibition at therapeutic concentrations by the AT(1) receptor antagonist losartan. *Br J Pharmacol* **134**, 1385-92.
- KHASAWNEH, F. T., HUANG, J. S., MIR, F., SRINIVASAN, S., TIRUPATHI, C. & LE BRETON, G. C. 2008. Characterization of isoprostane signaling: evidence for a unique coordination profile of 8-iso-PGF(2 α) with the thromboxane A(2) receptor, and activation of a separate cAMP-dependent inhibitory pathway in human platelets. *Biochem Pharmacol* **75**, 2301-15.
- KIKKAWA, U., ASE, K., OGITA, K. & NISHIZUKA, Y. 1986. [The role of protein kinase C in cell surface signal transduction and tumor promotion]. *Gan To Kagaku Ryoho* **13**, 861-9.
- MAGRANE M. AND THE UNIPROT CONSORTIUM. 2011. *UniProt Knowledgebase: a hub of integrated protein data* [Online]. UniProt. Available: <http://www.uniprot.org/uniprot/P21731> [Accessed November 20 2013].
- MARTIN, W. 1984. The combined role of atheroma, cholesterol, platelets, the endothelium and fibrin in heart attacks and strokes. *Med Hypotheses* **15**, 305-22.
- MONTUSCHI, P., BARNES, P. J. & ROBERTS, L. J., 2ND 2004. Isoprostanes: markers and mediators of oxidative stress. *Faseb j* **18**, 1791-800.
- MOREIRA, I. S. 2014. Structural features of the G-protein/GPCR interactions. *Biochim Biophys Acta* **1840**, 16-33.
- MORINELLI, T. A. & HALUSHKA, P. V. 1991. Thromboxane-A(2)/prostaglandin-H(2) receptors Characterization and antagonism. *Trends Cardiovasc Med* **1**, 157-61.
- MOUSA, S. A. 1999. Antiplatelet therapies: from aspirin to GPIIb/IIIa-receptor antagonists and beyond. *Drug Discov Today* **4**, 552-561.
- NAKAHATA, N. 2008. Thromboxane A2: physiology/pathophysiology, cellular signal transduction and pharmacology. *Pharmacol Ther* **118**, 18-35.
- NISHIMURA, A., KITANO, K., TAKASAKI, J., TANIGUCHI, M., MIZUNO, N., TAGO, K., HAKOSHIMA, T. & ITOH, H. 2010. Structural basis for the specific inhibition of heterotrimeric Gq protein by a small molecule. *Proc Natl Acad Sci U S A* **107**, 13666-71.

- OFFERMANN, S., LAUGWITZ, K. L., SPICHER, K. & SCHULTZ, G. 1994. G proteins of the G12 family are activated via thromboxane A2 and thrombin receptors in human platelets. *Proc Natl Acad Sci U S A*, 91, 504-8.
- PIRMOHAMED, M., JAMES, S., MEAKIN, S., GREEN, C., SCOTT, A. K., WALLEY, T. J., FARRAR, K., PARK, B. K. & BRECKENRIDGE, A. M. 2004. Adverse drug reactions as cause of admission to hospital: prospective analysis of 18 820 patients. *BMJ*, 329, 15-9.
- PUSCH, G., FEHER, G., KOTAL, K., TIBOLD, A., GASZTONYI, B., FEHER, A., PAPP, E., LUPKOVICS, G. & SZAPARY, L. 2008. Aspirin resistance: focus on clinical endpoints. *J Cardiovasc Pharmacol*, 52, 475-84.
- RUAN, C.Y., Zhou, W., Chan, C.H. 2011. Regulation of Smooth Muscle Contraction by the Epithelium: Role of Prostaglandins. *Physiology*, 26(3), 156-170.
- RASMUSSEN, S. G., DEVREE, B. T., ZOU, Y., KRUSE, A. C., CHUNG, K. Y., KOBILKA, T. S., THIAN, F. S., CHAE, P. S., PARDON, E., CALINSKI, D., MATHIESEN, J. M., SHAH, S. T., LYONS, J. A., CAFFREY, M., GELLMAN, S. H., STEYAERT, J., SKINIOTIS, G., WEIS, W. I., SUNAHARA, R. K. & KOBILKA, B. K. 2011. Crystal structure of the beta2 adrenergic receptor-Gs protein complex. *Nature*, 477, 549-55.
- ROTH, G. J., STANFORD, N., JACOBS, J. W. & MAJERUS, P. W. 1977. Acetylation of prostaglandin synthetase by aspirin. Purification and properties of the acetylated protein from sheep vesicular gland. *Biochemistry*, 16, 4244-8.
- SHIAU, Y.-F., HU, C.-J. & CHIUEH, C. C. 2012. Preventive Effectiveness of Aspirin on Recurrent Stroke. *Journal of Experimental & Clinical Medicine*, 4, 203-208.
- SNAPINN, S. M. 2000. Noninferiority trials. *Current Controlled Trials in Cardiovascular Medicine*, 1, 19-21.
- SPURNEY, R. F., BERNSTEIN, R. J., RUIZ, P., PISETSKY, D. S. & COFFMAN, T. M. 1991. Physiologic role for enhanced renal thromboxane production in murine lupus nephritis. *Prostaglandins*, 42, 15-28.
- TING, H. J., MURAD, J. P., ESPINOSA, E. V. & KHASAWNEH, F. T. 2012. Thromboxane A2 receptor: biology and function of a peculiar receptor that remains resistant for therapeutic targeting. *J Cardiovasc Pharmacol Ther*, 17, 248-59.